

I'm not a bot



Beta-lactam antibiotics have been a cornerstone in combating bacterial infections, offering solutions for many common and severe ailments. These drugs are valued for their broad-spectrum efficacy and relatively low toxicity compared to other antibiotic classes. However, antibiotic resistance poses challenges to their continued effectiveness. This article explores various aspects of beta-lactam antibiotics, including their mechanisms, types, and how bacteria develop resistance, providing insights into current medical practices and future considerations.

Mechanism of Action Beta-lactams target the bacterial cell wall, essential for maintaining cell integrity and shape. The cell wall is primarily composed of peptidoglycan, a polymer that provides mechanical strength. Beta-lactams interfere with the synthesis of this component by binding to penicillin-binding proteins (PBPs), enzymes involved in the final stages of peptidoglycan assembly. This binding inhibits the cross-linking of peptidoglycan strands, leading to a weakened cell wall. As the bacterial cell attempts to grow and divide, the compromised cell wall cannot withstand the internal osmotic pressure, resulting in cell lysis and death. This mechanism is particularly effective against actively dividing bacteria, as they are constantly remodeling their cell walls. The specificity of beta-lactams for bacterial cells, as opposed to human cells, accounts for their relatively low toxicity, making them a preferred choice in many clinical settings. The effectiveness of beta-lactam antibiotics is influenced by their ability to reach and bind to PBPs, which can vary among different bacterial species. Some bacteria possess multiple PBPs with varying affinities for beta-lactams, affecting their susceptibility. Additionally, the presence of efflux pumps and other resistance mechanisms can further limit the drug's effectiveness. The diversity of beta-lactams encompasses a diverse group of compounds, each with unique structural features and antibacterial spectra. These variations allow for targeted treatment of a wide range of bacterial infections. The primary classes include penicillins, cephalosporins, carbapenems, and monobactams, each offering distinct advantages and applications in clinical practice. Penicillins Penicillins are the earliest discovered beta-lactam antibiotics, with penicillin G being the first to be used clinically. They are primarily effective against Gram-positive bacteria, such as *Streptococcus* and *Staphylococcus* species. Over time, various derivatives have been developed to enhance their spectrum and stability, including amoxicillin and ampicillin, which are effective against some Gram-negative bacteria. Resistance to penicillins has emerged, often due to the production of beta-lactamase enzymes by bacteria, which hydrolyze the beta-lactam ring, rendering the antibiotic ineffective. To counteract this, beta-lactamase inhibitors like clavulanic acid are sometimes combined with penicillins to restore their efficacy. Cephalosporins Cephalosporins are a broad class of beta-lactam antibiotics developed over several generations, each with expanded activity against a wider range of bacteria. The first-generation cephalosporins, such as cephalexin, are effective primarily against Gram-positive bacteria. Subsequent generations, including cefuroxime and ceftazidime, have increased efficacy against Gram-negative organisms and are often used to treat infections like pneumonia and urinary tract infections. The structural modifications in cephalosporins enhance their resistance to beta-lactamases, making them a valuable option in cases where penicillin resistance is a concern. However, the emergence of extended-spectrum beta-lactamases (ESBLs) poses a challenge, as these enzymes can degrade many cephalosporins, necessitating careful selection and use in clinical settings. Carbapenems Carbapenems, such as imipenem and meropenem, are among the most potent beta-lactam antibiotics, known for their broad-spectrum activity against both Gram-positive and Gram-negative bacteria. They are particularly effective against multidrug-resistant strains, including those causing hospital-acquired infections. Their use is typically reserved for severe infections due to their potential for adverse effects, such as seizures at high doses. Monobactams Monobactams, such as aztreonam, are unique class members of beta-lactam antibiotics characterized by a single beta-lactam ring. They are specifically effective against aerobic Gram-negative bacteria, including *Pseudomonas aeruginosa*, making them useful in treating infections where Gram-negative pathogens are predominant. Monobactams are particularly valuable for patients with allergies to other beta-lactams, as they have a low cross-reactivity with penicillins and cephalosporins. While monobactams are resistant to some beta-lactamases, they are susceptible to others, such as metallo-beta-lactamases, which can limit their effectiveness. Their narrow spectrum of activity necessitates careful consideration of the infecting organism and susceptibility patterns when selecting monobactams for treatment. Resistance Mechanisms The emergence of resistance to beta-lactam antibiotics is a growing concern, driven by the adaptive capabilities of bacteria to survive in the presence of these drugs. One primary mechanism by which bacteria develop resistance is through the modification of target sites. Bacteria can alter or acquire mutations in the genes encoding penicillin-binding proteins (PBPs), reducing the binding affinity of beta-lactams. This alteration allows the bacteria to continue synthesizing their cell walls despite the presence of the antibiotic, thereby circumventing its intended effect. Additionally, efflux pump systems in bacteria contribute significantly to resistance. These pumps actively expel antibiotics from the bacterial cell, decreasing the intracellular concentration of the drug and thus its efficacy. Efflux pumps also play a role in resistance to other antimicrobial agents, highlighting their importance in bacterial survival. Another key mechanism of resistance involves the production of beta-lactamase enzymes. These enzymes catalyze the hydrolysis of the beta-lactam ring, effectively destroying the antibiotic before it can exert its effect. The acquisition of mobile genetic elements, such as plasmids that carry beta-lactamase genes, can spread resistance across different bacterial species, exacerbating the challenge of controlling resistant infections. Class of Broad-Spectrum Antibiotics β -Lactam Antibiotic Drug Class Core Structure of Penicillins (top) and Cephalosporins (bottom) The 2 most common groups of β -lactam antibiotics β -lactam ring in red. Class identifiers: UseBacterial InfectionATC codeJ01CBiological targetPenicillin binding proteinExternal linksMeSHD04790Legal statusIn Wikidata β -Lactam antibiotics (beta-lactam antibiotics) are antibiotics that contain a β -lactam ring in their chemical structure. This includes penicillin derivatives (penams), cephalosporins and cephamycins (cephems), monobactams, carbapenems[1] and carbacephem.[2] Most β -lactam antibiotics work by inhibiting cell wall biosynthesis in the bacterial organism and are the most widely used group of antibiotics. Until 2003, when measured by sales, more than half of all commercially available antibiotics in use were β -lactam compounds.[3] The first β -lactam antibiotic discovered, penicillin, was isolated from a strain of *Penicillium rubens* (named as *Penicillium notatum* at the time).[4][5] Bacteria often develop resistance to β -lactam antibiotics by synthesizing a β -lactamase, an enzyme that attacks the β -lactam ring. To overcome this resistance, β -lactam antibiotics can be given with β -lactamase inhibitors such as clavulanic acid. [6] β -lactam antibiotics are indicated for the prevention and treatment of bacterial infections caused by susceptible organisms. At first, β -lactam antibiotics were mainly active only against gram-positive bacteria, yet the recent development of broad-spectrum β -lactam antibiotics active against various gram-negative organisms has increased their utility. β -lactams are categorized into two main classes based on the core structure of the molecule: penicillins and cephalosporins. Both classes share a common four-membered β -lactam ring fused to a five-membered thiazolidine ring. The difference lies in the side chain attached to the nitrogen atom of the β -lactam ring. Penicillins have a phenylacetamido side chain, while cephalosporins have a dihydrothiazolidine side chain. This structural similarity between β -lactam antibiotics and α -amino- γ -alanine facilitates their binding to the active site of PBPs. The β -lactam nucleus of the molecule irreversibly binds to (acylates) the Ser62 residue of the PBP active site. This irreversible inhibition of the PBPs prevents the final crosslinking (transpeptidation) of the nascent peptidoglycan layer, disrupting cell wall synthesis.[13] β -Lactam antibiotics block not only the division of bacteria, including cyanobacteria, but also the division of cyanelles, the photosynthetic organelles of the glaucophytes, and the division of chloroplasts of bryophytes. In contrast, they have no effect on the plastids of the highly developed vascular plants. This is supporting the endosymbiotic theory and indicates an evolution of plastid division in land plants.[14] Under normal circumstances, peptidoglycan precursors signal a reorganisation of the bacterial cell wall and, as a consequence, trigger the activation of autolytic cell wall hydrolases. Inhibition of cross-linkage by β -lactams causes a build-up of peptidoglycan precursors, which triggers the digestion of existing peptidoglycan by autolytic hydrolases without the production of new peptidoglycan. As a result, the bactericidal action of β -lactam antibiotics is further enhanced.[citation needed] Another possibility that has been proposed to account for much of the cytotoxicity of β -lactams focuses on the oxidation of the guanine nucleotide in the bacterial nucleotide pool.[15] The incorporation of oxidized guanine nucleotide into DNA could cause cytotoxicity. Bacterial cytotoxicity could arise from incomplete repair of closely spaced 8-oxo-2'-deoxyguanosine lesions in the DNA resulting in double-strand breaks.[15] It is also possible that the structural features of β -lactam antibiotics have been selected for their cytotoxicity. The pyrimidine parameter, h, and is related to the height in angstroms of the pyramid formed by the nitrogen atom of the β -lactam as the apex and the three adjacent carbon atoms as base [17]. The β -lactam nucleus is a bicyclic system consisting of a four-membered β -lactam ring fused to a five-membered thiazolidine ring. The β -lactam ring is responsible for conferring resistance to β -lactam antibiotics. The hydrolysis of the β -lactam ring by β -lactamase enzymes renders the antibiotic inactive, thereby allowing the bacteria to survive in the presence of the antibiotic.[28] Some β -lactams, such as piperacillin and ticarcillin, are modified with a meropenem concentration dropped to 90% of the initial concentration at 7.4 hours at 22°C and 5.7 hours at 33°C, indicating degradation over time.[29] As a response to the use of β -lactams to control bacterial infections, some bacteria have evolved penicillin binding proteins with novel structures. β -Lactam antibiotics cannot bind as effectively to these altered PBPs, and, as a result, the β -lactams are less effective at disrupting cell wall synthesis. Notable examples of this mode of resistance include methicillin-resistant *Staphylococcus aureus* (MRSA)[32] and penicillin-resistant *Streptococcus pneumoniae*. Altered PBPs do not necessarily rule out all treatment options with β -lactam antibiotics. [medical citation needed] See also: List of β -lactam antibiotics The β -lactam core structures. (A) A penam. (B) A carbapenam. (C) An oxapenam. (D) A penem. (E) A carbapenem. (F) A monobactam. (G) A cephem. (H) A carbacephem. (I) An oxacephem. β -Lactams are classified according to their core ring structures.[33] β -Lactams fused to saturated five-membered rings: β -lactams containing thiazolidine rings are named penams. β -lactams containing pyrrolidine rings are named carbapenams. β -lactams fused to unsaturated five-membered rings: β -lactams fused to unsaturated five-membered rings are named oxapenams or clavams. β -lactams fused to unsaturated six-membered rings: β -lactams containing 2,3-dihydrothiazole rings are named penems. β -lactams containing 2,3-dihydro-1H-pyrrole rings are named carbapenams. β -lactams fused to unsaturated six-membered rings: β -lactams containing 3,6-dihydro-2H-1,3-thiazine rings are named cephs. β -lactams containing 1,2,3,4-tetrahydropyridine rings are named carbacephems. β -lactams containing 3,6-dihydro-2H-1,3-oxazine rings are named oxacephems. β -lactams not fused to any other ring are named monobactams. By convention, the bicyclic β -lactams are numbered starting with the position occupied by sulfur in the penams and cephs, regardless of which atom it is in a given class. That is, position 1 is always adjacent to the β -carbon of β -lactam ring. The numbering continues clockwise around the β -lactam ring. In carbapenams, the β -lactam ring is fused second in sequence.[citation needed] The biosynthesis of the β -lactam ring of tetracycline involves the condensation of L-proline and L-glutamine. The closure of the β -lactam ring in the other monobactams, such as sulfazecin and the nocardimides, may involve a third mechanism involving an amino acid derivative. The β -lactam ring is fused second in sequence.[citation needed] The biosynthesis of the β -lactam ring of tetracycline involves the condensation of L-proline and L-glutamine. 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per species (Georgopapadakou and Liu 1980). The inhibition of bacterial peptidoglycan transpeptidation was described mechanistically by Tipper and Stromberg (1965), who noted a structural similarity of penicillin G to the terminal d-Ala-d-Ala dipeptide of the nascent peptidoglycan in the dividing bacterial cell. This mechanism is now known to involve binding of penicillin, or another β-lactam, to an active site serine found in all functional PBPs (Georgopapadakou et al. 1977). The resulting inactive acyl enzyme may then slowly hydrolyze the antibiotic to form a microbiologically inactive entity (Frère and Joris 1985). In addition to these functionalities, recent work has shown the binding of selected β-lactams, such as cefaroline, to an allosteric site in PBP2a from *Staphylococcus aureus*, resulting in an increased sensitization of the organism to the antibiotic (Otero et al. 2013; Gonzales et al. 2015). PBPs may be divided into classes according to molecular mass (Coffin and Ghuysen 1998; Massova and Mobashery 1998), with low-molecular-mass PBPs serving mainly as monofunctional d-Ala-d-Ala carboxypeptidases. High-molecular-mass PBPs have been divided into two subclasses, one of which (class A) includes bifunctional enzymes with both a transpeptidase and a transglycosylase domain, and the second of which (class B) encompasses d-Ala-d-Ala-dependent transpeptidases. At least one PBP is deemed to be essential in each species, with a unique specificity for β-lactam binding that varies among each species and each β-lactam class (Curtis et al. 1979; Georgopapadakou and Liu 1980). In Gram-negative bacteria, essential PBPs include the high-molecular-weight PBPs 1a and 1b that are involved in cell lysis, PBP2, the inhibition of which results in a cessation of cell division and the formation of spherical cells, and PBP3 for which inhibition arrests cell division, resulting in filamentation. Cell death may occur as a result of inhibiting one or more of these PBPs (Spratt 1977, 1983). The roles of PBPs in Gram-positive bacteria and *Mycobacterium tuberculosis* are discussed in detail in Fisher and Mobashery (2016). Penicillin G (benzylpenicillin) was the first β-lactam to be used clinically, most frequently to treat streptococcal infections for which it had high potency (Rammelkamp and Keefer 1943; Hirsch and Dowling 1946). Another naturally occurring penicillin, penicillin V (phenoxymethylpenicillin), in an oral formulation is still used therapeutically and prophylactically for mild to moderate infections caused by susceptible *Streptococcus* spp., including use in pediatric patients (Pottegard et al. 2015). However, the selection of penicillin-resistant penicillinase-producing staphylococci in patients treated with penicillin G led to decreased use of this agent, and prompted the search for more penicillins with greater stability to the staphylococcal β-lactamases (Kirby 1944, 1945; Medeiros 1984). A list of historically important and clinically useful penicillins is provided in Table 2. Among the penicillinase-stable penicillins of clinical significance are methicillin, oxacillin, cloxacillin, and nafcillin, with the latter suggested as the β-lactam of choice for skin infections, catheter infections, and bacteremia caused by methicillin-susceptible *S. aureus* (Bamberger and Boyd 2005). All were used primarily for staphylococcal infections until the emergence of methicillin-resistant *S. aureus* (MRSA) in 1979–1980 (Hemmer et al. 1979; Saroglou et al. 1980). Penicillins of current and historical utility Penicillins with improved activity against Gram-negative pathogens included the orally bioavailable ampicillin and amoxicillin, both of which were introduced in the 1970s. These agents were initially used for the treatment of infections caused by Enterobacteriaceae and did not effectively inhibit the growth of *Pseudomonas aeruginosa*, which became more of a concern during the late 1970s. Carbenicillin was the first antipseudomonal penicillin to be introduced, but lacked stability to β-lactamase hydrolysis and was less potent than piperacillin or ticarcillin, later antipseudomonal penicillins. These latter drugs were considered to be potent broad-spectrum penicillins that included penicillin-susceptible staphylococci, enteric bacteria, anaerobes, and *P. aeruginosa* in their spectrum of activity. They were used extensively to treat serious nosocomial infections, especially when combined with a β-lactamase inhibitor (see below). Two parenteral penicillins with unusual chemical structures, mecillinam and temocillin (Table 2), were introduced to treat infections caused by enteric bacteria before the global emergence of extended-spectrum β-lactamases (ESBLs) in the late 1980s. Mecillinam (also known as amdinocillin), with a 6-β-amidino side chain, is a narrow-spectrum β-lactam that binds exclusively to PBP2 in enteric bacteria (Curtis et al. 1979). Because of this specificity, it shows synergy in vitro in combination with other β-lactams that bind to PBPs 1a/1b and/or PBP3 in Gram-negative bacteria (Hanberger et al. 1991), thus decreasing the possibility that a point mutation in a single PBP would lead to resistance (Hickman et al. 2014). Temocillin, the 6-α-methoxypenicillin analog of ticarcillin, had greater stability than ticarcillin to hydrolysis by serine β-lactamases, but lost antibacterial activity against Gram-positive bacteria, anaerobic Gram-negative pathogens, and some enteric bacteria that included the important pathogens *Enterobacter* spp. and *Serratia marcescens* (Martinez-Beltran et al. 1985). Mecillinam and temocillin are currently enjoying a resurgence in interest owing to their stability to many ESBLs (Livermore et al. 2006; Rodríguez-Villalobos et al. 2006), often resulting in greater than 90% susceptibility when tested against many contemporary ESBL-producing Enterobacteriaceae (Giske 2015; Zykov et al. 2016). Because increasing numbers of β-lactamases have compromised the use of penicillins as single agents (Bush 2013), there is currently limited therapeutic use of the penicillins as monotherapy. Ampicillin, amoxicillin, piperacillin, and ticarcillin have continued to be useful, primarily as a result of their combination with an appropriate β-lactamase inhibitor (see below). However, even ampicillin, amoxicillin, penicillin G, and penicillin V are still active as monotherapy against Group A streptococci, and *Treponema pallidum*, two of the few bacterial species that do not produce β-lactamases (Schaar et al. 2014). During the 1950s, the discovery of the naturally occurring penicillinase-stable cephalosporin C opened a new pathway to the development of hundreds of novel cephalosporins (Newton and Abraham 1956; Abraham 1987) to treat infections caused by the major penicillinase-producing pathogen of medical interest at that time, *S. aureus*. Dozens of cephalosporins were introduced into clinical practice (Abraham 1987), either as parenteral or oral agents. The molecules exhibited antibacterial activity with MICs often ≤4 μg/mL against not only staphylococci, but also *Streptococcus pneumoniae* and non-β-lactamase-producing enteric bacteria. The parenteral agents were generally eightfold more potent than the oral agents that were used in some cases to replace oral penicillins in penicillin-allergic patients. The early cephalosporins, for example, those in the cephalosporin I subclass (Bryskier et al. 1994) introduced before 1980, were labile to hydrolysis by many β-lactamases that emerged following their introduction into clinical practice, so that only a few of the early molecules remain in use (see Table 3), primarily to treat mild to moderate skin infections caused by methicillin-susceptible *S. aureus* (MSSA) (Giordano et al. 2006). Cefazolin with high biliary concentrations is still used for surgical prophylaxis and for treatment of abdominal infections (Sudo et al. 2014) and is effective as empiric therapy in 80% of Japanese children with their first upper urinary tract infection (Abe et al. 2016). Cephalosporins of current clinical utility or of historical interest When the TEM-1 penicillinase began to appear on transmissible plasmids in *Neisseria gonorrhoeae* (Ashford et al. 1976) and *Haemophilus influenzae* (Gunn et al. 1974; Khan et al. 1974), it was quickly recognized that the penicillins and cephalosporins in medical use were becoming ineffective, not only in treating those TEM-1-producing organisms, but also for the enteric bacteria and *P. aeruginosa* that could all acquire this enzyme. Another surge of synthetic activity in the pharmaceutical industry provided both oral and parenteral cephalosporins with stability to this common enzyme. These agents tended to have decreased potency against the staphylococci, but gained antibacterial activity against Gram-negative pathogens. Cefuroxime, dosed parenterally or orally as the axetil ester, was the only member of the cephalosporin II class (Bryskier et al. 1994) with both oral and systemic dosage forms, but its stability to β-lactamase hydrolysis was diminished compared to later oral cephalosporins (Jacoby and Carreras 1990). As seen with cefuroxime, acceptable oral bioavailability of cefpodoxime required esterification through addition of a proxetil group to attain sufficient absorption for efficacy (Bryskier and Belfiglio 1999). Of the oral agents approved after 1983 in Table 3, cefdinir was generally more stable to hydrolysis, not only to the original TEM enzyme, but also to the AmpC cephalosporinases that are produced at a basal level in many enteric bacteria and *P. aeruginosa* (Payne and Amyes 1993; Labia and Morand 1994). Among the parenteral agents introduced in the 1980s were the cephamycin cefoxitin, and cephalosporins in the cephalosporin III and cephalosporin IV subclasses (Bryskier et al. 1994), which continue to serve as important antibiotics for the treatment of serious infections caused by Gram-negative pathogens. The novel oxacephem moxalactam, or latamoxef, which had similar antimicrobial activity to the cephalosporin III/IV subclasses, has exquisite stability to hydrolysis by β-lactamases (Sato et al. 2015), but was not a highly successful antibiotic owing, in part, to a relatively high frequency of bleeding in patients treated with this drug (Brown et al. 1996). The cephamycin cefoxitin is notable for its characteristic 7-methoxy side chain that confers stability to the TEM-type β-lactamases, including ESBLs. It has useful antibacterial activity against MSSA and enteric bacteria that do not produce high levels of AmpC cephalosporinases (Jacoby and Han 1996). Cefotaxime, cefoperazone, ceftriaxone, and ceftazidime, designated as subclass cephalosporin III, and cefepime in the cephalosporin IV subclass, are also known as expanded-spectrum cephalosporins with increased hydrolytic stability to the common penicillinases, SHV-1 and TEM-1 β-lactamase (Martinez-Martinez et al. 1996). These agents have diminished activity against staphylococci and enterococci compared to earlier cephalosporins, but have more potent activity against Gram-negative organisms. Cefepime tends to have lower MICs against enteric bacteria than the other expanded-spectrum cephalosporins, attributed to greater penetration through the OmpF outer-membrane porin protein (Nikaido et al. 1990; Bellido et al. 1991). Cefotaxime and ceftriaxone are often used to treat susceptible streptococcal infections; all can be used to treat serious infections caused by enteric bacteria if the organisms test susceptible. Notably, ceftazidime and cefepime have maintained their observed activity against *P. aeruginosa*, with recent susceptibility rates exceeding 80% (Sader et al. 2015). A liability of the expanded-spectrum cephalosporins, however, began to emerge only a few years after the introduction of cefotaxime, when the ESBLs were identified with the ability to hydrolyze all of the β-lactams, with the exception of the carbapenems. These enzymes, in addition to both serine and metallo-carbapenemases, have severely compromised the activity of almost all penicillins and cephalosporins, necessitating the development of combination therapy with other β-lactams, β-lactamase inhibitors, or antibiotics from other classes. Cefzolozane, recently approved in combination with tazobactam for the treatment of complicated urinary tract infections and complicated intraabdominal infections, shows potent antipseudomonal activity, and includes activity against enteric bacteria that produce some ESBLs (Zhanel et al. 2014), particularly CTX-M-producing isolates (Estabrook et al. 2014). Another recent addition to the cephalosporin family is the siderophore-substituted cephalosporin S-849266 with a catechol in the 3-position, thus allowing the molecule to enter the cells via an iron transport mechanism (Kohira et al. 2015). In addition to increased penetrability, the cephalosporin is stable to hydrolysis by many carbapenemases, resulting in activity against many β-lactam-resistant enteric bacteria (Kohira et al. 2015). In the mid-1990s, reports began to emerge describing cephalosporins with MICs